



6-Acylamino-penam Derivatives: Synthesis and Inhibition of Cathepsins B, L, K, and S

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Received 5 July 2002; accepted 26 August 2002

Abstract—The synthesis of a new series of 6-acylamino penam derivatives and their inhibition of cysteine proteases cathepsins B, L, K, and S is described. The 6-acylamino-penam sulfone compounds showed excellent cathepsin L, K, and S inhibition activity with IC₅₀ values in the nanomolar and subnanomolar range.

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The cysteine proteases cathepsin B, L, K, and S may be involved in diseases such as osteoporosis, cancer metastasis, rheumatoid arthritis, and infectious diseases. These proteases are implicated as an important targets for the development of inhibitors as potential therapeutic agents. Several types of chemical functionalities served as the central pharmacophore for cysteine proteases inhibitors, such as aldehydes, nitriles, α -ketocarbonyl compounds, halomethyl ketones, acyloxymethyl ketone, epoxide, vinyl sulfones, cyclopropenone and cyclohexanone. 3,4

$$Ph \longrightarrow O \longrightarrow N \longrightarrow H \longrightarrow N \longrightarrow N \longrightarrow N$$

We have reported on the rational design and synthesis of a series of 6-substituted-4-oxa-1-azabicyclo[3,2,0]heptan-7-one derivatives, 1, as cysteine protease inhibitors. Although potent inhibition of cathepsin L and K was achieved with IC_{50} values in the nanomolar range (for compound 1; IC_{50} is 4 and 5 nM for cathepsin L and K, respectively), one concern with such inhibitors was their potential toxicity, since the 4-oxa-1-azabicyclo[3,2,0]

heptan-7-one skeleton is reported to show cytotoxic activity. 6 The alternative structure related to 4-oxa-1-azabicyclo[3,2,0]heptan-7-one skeleton is the penam which is a well known pharmacophore for antibiotics and is widely used in clinic.⁷ Therefore, a new series of 6-acylaminopenam derivatives was synthesized and their inhibitions with cathepsins B, L, K, and S were evaluated. The synthesis of 6-acylamino-penam compounds is outlined in Scheme 1. The intermediate 4-acetoxy-3-acylamino-azetdin-2-one 2 was prepared according to our previous report⁸ from 6-aminopenicillanic acid. Substitution of the acetate 2 with mercaptoethanol in the presence of NaOH gave 4-hydroxyethylthio-azetidin-2-one. Treatment of the hydroxy compound with toluenesulfonyl chloride under standard conditions gave toluenesulfonyloxyethylthioazetidin-2-one, which was converted to 4-bromoethylthioazetidin-2-one, 3, with lithium bromide. Without further purification, cyclization of 4-bromoethylthio-azetidin-2one 3 with base gave two optically pure isomers of the penam derivatives 4 and 5 in high yield after silica gel column chromatography. A careful and controlled oxidation of compound 4 with hydrogen peroxide in acetic acid⁹ gave a mixture of 4β - and 4α -penam sulfoxides 6 in ratio of 3:1.10 Complete oxidation of 4 or 5 with KMnO₄ resulted in penam sulfones 7 or 8, respectively. For comparison, two cephem sulfone derivatives 9 and 10 were also prepared by a similar method as described above. Using a similar method, compounds 11–15 were prepared as well, where phenylalanine was replaced by cyclohexylalanine, 2-naphthylalanine, 2-thiophenalanine, 3-fluorophenylalanine, or 3-pyridylalanine, respectively.

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Scheme 1. Reagents and conditions: (a) $HS(CH_2)_2OH$, 1 equiv NaOH, THF/H_2O , rt 1 h; (b) TsCl, pyridine, 5 °C, 6 h; (c) LiBr, HMPA, 60 °C, 2 h; (3) 45% for three step reaction; (d) K_2CO_3 , DMSO, rt overnight; (4) 65%; (5) 25%; (e) 1.5 equiv H_2O_2 , 4 equiv AcOH, DCM, rt 2 days; (6) 50%; (f) $KMnO_4$, AcOH, H_2O , rt, 1 h; (6) 65%; (7) 45%.

The inhibitory activities of these compounds with cathepsin B, L, K, and S were determined according to the procedure described in the literature¹¹ by using Cbz-Phe-Arg-AMC for cathepsin B, L, and K and Cbz-Val-Val-Arg-AMC for cathepsin S. The inhibitory activities of the compounds are shown in Table 1. The 6-acylamino-penam sulfone derivatives are most potent inhibitors of cathepsin L, K, and S with IC₅₀ in sub-

Table 1. In vitro inhibitory activity of azetidin-2-one derivatives with cysteine proteases

Compd	IC_{50} (μM)			
	Cath B	Cath L	CathK	Cath S
4	> 50	6.68	> 2.5	n.d.a
6	16.4	0.068	0.035	0.07
7	0.35	0.0014	0.0005	0.01
8	6.01	0.0096	0.0075	0.21
9	> 50	10.6	> 2.5	> 2.5
10	> 50	> 50	> 2.5	> 2.5
11	0.43	0.0007	0.0076	0.0007
12	0.39	0.003	0.4	0.003
13	0.43	0.003	0.0018	0.0046
14	1	0.003	0.006	0.008
15	1.3	0.0017	0.0025	0.058

an.d., not determined.

nanomolar or low nanomolar range. It appears that activity correlates strongly with an oxidation state of the penam ring (compare the activity of compound 4 with compounds 6 and 7). The sulfones are more active than sulfoxide and sulphide, may be due to the increased chemical reactivity of azetidin-2-one skelton. The 5S stereo isomer compound 7 is preferred for better cysteine proteases inhibitory activity over the 5R stereo isomer compound 8 as in our previous finding.⁵ In contrast, six-membered ring cepham sulfones do not exhibit any inhibitory activity. Different aromatic and heterocyclic substitutions at R, mimicking presumably P2 portion revealed lack of selectivity (11, 13, 14, and 15), with exception of a naphthyl substituent which is not tolerated well with cathepsin K, but better with other cathepsins (12). Penam sulfoxides and sulfides are fully reversible inhibitors, while sulfones are slow reversible and time dependent inhibitors.

References and Notes

1. Delaisse, J. M.; Ledent, P.; Vaes, G. *Biochem. J.* 1991, 279, 167. Thomson, S. K.; Halbert, S. M.; Bossard, M. *J. Proc. Natl. Acad. Sci. U.S.A.* 1997, 94, 14249. Maciewicz, R. A.; Van der Stapper, J. W. J.; Paraskewa, C.; Williams, A. C.; Hague, A. *Biochem. Soc. Trans.* 1991, 19, 362. Esser, R. E.;

- Angelo, R. A.; Murphey, M. B.; Watts, L. M.; Thornburg, L. P.; Palmer, J. T.; Talhouk, J. W.; Smith, R. E. Arthritis Rheum. 1994, 37, 236. Semenov, A.; Olson, J. E.; Rosental, P. J. Antimicrobiol. Agents and Chemother. 1998, 42, 2254. Engel, C.; Doyle, P.; Hsieh, I.; McKerrow, J. H. J. Exp. Med. 1998, 188, 725.
- 2. Chapman, H. A.; Riese, R. J.; Shi, G. P. Annu. Rev. Physiol. 1997, 59, 63. Smith, W. W.; Abdel-Meguid, S. S. Exp. Opin. Ther. Pat. 1999, 9, 683. Henkin, J. Annu. Rep. Med. Chem. 1993, 28, 151. Michaud, S.; Gour, B. J. Exp. Opin. Ther. Pat. 1998, 8, 645. Elliott, E.; Sloane, B. F. Exp. Opin. Ther. Pat. 1996, 6, 12.
- 3. Reviews on cysteine proteases inhibitor: Otto, H.; Schirmeister, T. Chem. Rev. 1997, 133. Yamashita, D. S.; Dodds, R. A. Curr. Pharm. Des 2000, 6, 1. Leung, D.; Abbenante, G.; Fairlie, D. P. J. Med. Chem. 2000, 43, 305. Marquis, R. W. Annu. Rep. Med. Chem. 2000, 35, 309. Leung-Toung, R.; Li, W.; Tam, T. F.; Karimian, K. Curr. Med. Chem. 2002, 9, 979. 4. Conroy, J. L.; Sanders, T. C.; Seto, C. T. J. Am. Chem. Soc. 1997, 119, 4285. Conroy, J. L.; Abato, P.; Ghosh, M.; Austermuhle, M. I.; Kiefer, M. R.; Seto, C. T. Tetrahedron Lett. 1998, 39, 8253.

- 5. Zhou, N. E.; Guo, D.; Kaleta, J.; Purisima, E.; Menard, R.; Micetich, R. G.; Singh, R. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 3413
- Otani, T.; Oie, S.; Matsumoto, H.; Tempest, M.; Micetich,
 R. G.; Singh, R.; Yamashita, T. EP Patent 0 654 995 B1, 1993.
 Chem. Abstr. 1994, 121, 57497.
- 7. Newall, C. E., Hallam, P. D. In *Comprehensive Medicinal Chemistry*; Hansch, C., Sammes, P. J., Taylor, J. B., Eds.; Pergamon: Oxford 1990; p 605.
- 8. Zhou, N. E.; Guo, D.; Thomas, G.; Kaleta, J.; Micetich, R. G.; Singh, R. *Bioorg. Med. Chem. Lett.* Submitted for publication.
- 9. Micetich, R. G.; Singh, R.; Shaw, C. C. *J. Org. Chem.* **1986**, *51*, 1811. Micetich, R. G.; Singh, R.; Maiti, S. N. *Heterocycles* **1984**, *22*, 531.
- 10. The separation of isomers was not achieved by column chromatography due to close R_f and the ratio of isomers is determined based on C5 proton by NMR.
- 11. Barrett, A. J.; Kembhavi, A. A.; Brown, M. A.; Kirschke, H.; Knight, G.; Tamai, M.; Hanaka, K. *Biochem. J.* **1982**, *201*, 189.